Stereospecific Coupling Reaction for lnternucleotlde Methyl Phosphonothioate Linkage

Alexander V. Lebedev', Jason P. Rife, Howard W. Seligsohn, George R. Wenzinger, and Eric Wickstromt.

Department of Chemistry, University of South Florida, Tampa, Florida 33620, USA *Institute of Bioorganic Chemistry, Siberian Division of the Academy of Sciences of the USSR, Novosibirsk, 630090, USSR

t To whom correspondence should be addressed.

Abstract: The (Sp)- and (Rp)- diastereomers of 5'-dimethoxytritylthymidyl-(3'-5') methylphosphono-5'-thio-3'-acetylthymidine were prepared by the stereospecific reaction of 5'-deoxy-5'-iodo-3'-acetylthymidine with the (Sp)- or (Rp)- diastereomers of, 5'-dimethoxytritylthymidyl-3' methylphosphonothioate (Li+ salt) in DMF. At 50° C, the coupling reaction was complete within 6 hours.

Oligodeoxynucleotide derivatives with uncharged internucleotide groups, such as methylphosphonates¹ and alkyl phosphotriesters²⁻⁴, are useful as probes of nucleic acid interactions with proteins and nucleic acids, as diagnostic probes, and as therapeutic agents⁵⁻⁷. These derivatives penetrate cell membranes easily and are highly resistant to nucleases⁸⁻¹¹. However, this type of modification generates a chiral center at the phosphorus atoms, yielding nonequivalent diastereomeric oligodeoxynucleotide derivatives. Hybridization of the above oligodeoxynucleotide derivatives with complementary sequences of nucleic acids is highly dependent on the absolute configuration of the phosphorus centers¹²⁻¹⁶. Hence, stereoselective coupling reactions are being developed in several laboratories^{17,18}. In this paper we report a stereospecific synthesis of P-S-C(5') modified dideoxynucleoside methylphosphonate.

First, both (Rp)-and (Sp)-isomers of the key intermediate, 5'-dimethoxytritylthymidyl-3' methylphosphonothioate¹⁹ (Li⁺ salt) 4 were prepared according to scheme 1. 2-cyanoethylation of 1 was carried out in the presence of 4-(N,N-diethylamino)-pyridine (DMAP) and trifluoroacetic anhydride²⁰. The reaction was followed with $31P$ NMR spectroscopy; the chemical shifts of the reactants and products are given in scheme 1. The diastereomers of 3 were purified and separated by silica HPLC into two components designated **&fast** and 3-slow. The absolute configurations have not been determined. The cyanoethyl groups were removed with concentrated ammonium hydroxide: ethanol (1:2). The deprotected monomers were purified by silica HPLC. The ammonium cation was exchanged with Li+ by using a Dowex 50W x 2 column, yielding the products **4-fast** and 4-slow, whose structures were confirmed by ¹H NMR spectroscopy. 5'-deoxy-5'-iodo-3'acetylthymidine (5) was prepared by reaction of 3'-acetylthymidine and methyltriphenoxyphosphonium iodide in DMF²¹.

4 was coupled with either CH3l or 5 according to scheme 2, similarly to the synthesis of 5-thio

Table 1: Dependence of Coupling Rate on Concentration and Temperature.

For coupling with 5, solvent was DMF; for coupling with CH3I, solvent was acetonitrile, and 4-fast and 4-slow were ammonium salts.

oligodeoxynucleotides²². The coupling reaction with CH₃I was completed within 30 minutes. The products 6 were purified by silica HPLC with 2% EtOH in CHCl3, yielding either **6-fast (7** min.) or 6 slow (10 min.) whose structures were confirmed by ¹H NMR spectroscopy.

The rate of coupling of 4 with 5 was shown to be greatly dependent on temperature and concentration (Table.1). At low concentrations $([4] = [5] = 0.03$ M) and room temperature, the reactions required more than 10 days for completion. However, at room temperature and high concentrations $([4] = 0.14$ M, $[5] = 0.19$ M) the reaction was complete in less than four days. In contrast, at 50°C, the reactions were complete within 6 hrs., at all concentrations tested. The products were purified by silica HPLC with 3% EtOH in CHCl₃ yielding either 7-fast (8 min.) or 7-slow (10 min.), whose structures were confirmed with 1H NMR spectroscopy. Although 50-60% yields of 7 were obtained as dry powders after purification and precipitation, the $31P$ NMR and silica HPLC data implied that the coupling reaction was nearly quantitative. Even with the very long reaction time there were no indications of by-products in $31P$ NMR spectra.

In both coupling reactions, with either CHsl or 5,3-fast yielded only **6-fast** or **7-fast,** while 3 slow yielded only 6-slow or 7-slow, without racemization. Similar stereospecific coupling was obtained for the reaction of 5'-dimethoxytrityl-thymidyl-3'-methylthiophosphonate (prepared from separated diastereomers of the S-aryl derivative²³) and 5'-tosylthymidine in the presence of sodium iodide or tetrabutylammonium iodide as a catalyst¹⁸, but the rates of the reactions were at least 10 times slower.

The next goals of our research are to develop a scheme for solid phase synthesis following the same reaction pathway and to develop an alternative scheme that yields a methylphosphonate P-O- $C(5')$.

Acknowledgments: This work was supported by grants to E. W. from the National Cancer Institute, the Florida High Technology and Industry Council, and Genta. Inc.

References:

- 1. P.S.Miller, J.Yano, E.Yano, C.Carroll, K.Jayaraman, and P.O.P.Ts'o, Biochemistry, 18, 5134 (1979).
- 2. P.S.Miller, K.N.Fang, N.S.Kondo, and P.O.P.Ts'o, *J.Amer.Chem.Soc., 93, 6657* (1971).
- 3. W.J.Stec, **G.Zon,** K.A.Gallo, R.A.Byrd, B.Uznanski, and P.Guga, *Tetrahedron Lett., 26,2191* (1985).
- 4. H.M.Moody, M.H.P. van Genderen, L.H.Koole, H.J.M.Kocken, E.M.Meijer, and H.K.Buck, Nucleic *Acids Res. 17, 4769* (1989).
- 5. S.A.Noble, E.F.Fisher, and M.H.Caruthers, *Nucleic Acids Res., 12, 3367* (1984).
- 6. P.S. Miller, P.O.P. Ts'o, *Anti-cancer Dmg Design 2,* **117** (1987).
- 7. AR. van der Kml, J.N.M. Mel, AR. Stuitje, *Biotechniques, 6, 958,* (1988).
- 6. P.S.Miller, L.T.Braiterman, and P.O.P.Ts'o, *Biochemistry,* **16,** 1988 (1977).
- 9. G.G.Karpova, D.G.Knorre, A.S.Ryte, and L.E.Stephanovich, **FEBS Left, 122,21 (1980).**
- 10. C.H.Agris, K.R.Blake, P.S.Miller, M.P.Reddy, and P.O.P.Ts'o, *Biochemistry*, 25, 6268 (1986).
- 11. T.V.Abramova, V.V.Vlassov, A.V.Lebedev, and **A.S.Ryte,** *FEBS Lett., 236, 243* (1988).
- 12. P.S. Miller, N.D. Annan, K.B. McPartand, SM. Pulford, *Biochemistry, 21, 2507* (1962).
- 13. P.S.Miller, N.Dreon, S.M.Pulford, and K.B. MC Parland, *J.Bio/.Chem., 255, 9659* (1980).
- 14. M.F.Summers, G.Powell, W.Egan, R.A.Byrd, W.D.Wilson, and G.Zon, *NucleicAcids Res., 14, 7421* (1986).
- 15. M.F.Summers, C.Powell, K.Shinozuka, J.B.Regan, **G.Zon,** and W.D.Wilson, *Nucleic Acids Res.,* **15, 4915 (1987).**
- 16. R.L.Letsinger, S.A.Bach, J.S.Eadie, *Nucleic Asids Res., 14, 3467 (1986).*
- 17. Z. **J.** Lesnikowski, M. Jaworska, W.J. Stec, *Nucleic Acids Res.,* **16,** 11675 (1986).
- 18. W. K. D. Brfll, and M.H. Caruthers, *Nocleofides d Nucleosides, 8(5&S),* 1011 (1989).
- 19. W. Niewiarowski, **Z.** J. Lesnikowski, A. Wilk, P.Guga, A. Okruszek, B.Uznanski, and, W.Stec, *Acta Biochimica Poi., 34, 217* (1987).
- 20. A.V. Lebedev, G.R. Wenzinger, and E. Wickstrom, Tetrahedron Lett.,Sl, preceding paper (1990).
- 21. J.P.H. Verheyden, and J.G. Moffat, *J. Org. Chem., 35,2319* **(1970).**
- 22. **V.P.** Kumarev, V.S. Bogachev, V.F. Kobsev, V.L. Baranova, M.I. Rivkin, V.N. Rybakov, *Bioorganicheskaya Khimia (Russ.)* ,8, 1525 **(1982).**
- 23. W.K.D. Brill, and M.H. Caruthers, *Tetrahedron Left., 29, 1227* (1968).